

Ultra-Fast, One-Pot Synthesis of Nanofibers by Lewis Pair Polymerization-Induced Self-Assembly

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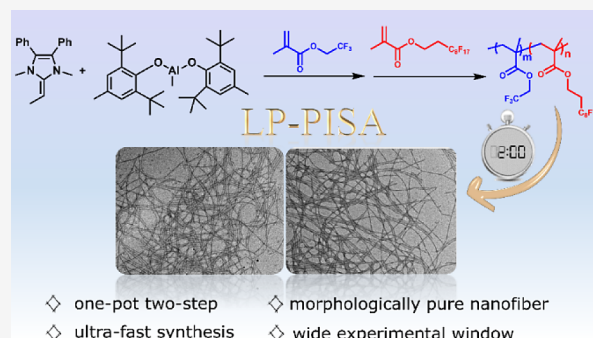


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ABSTRACT: Owing to their wide application potentials resulting from high aspect ratio, morphologically pure nanofibers attract intense attention but remain challenging to synthesize due to their narrow synthetic experimental window. Here, we employed strong nucleophilic *N*-heterocyclic olefin (NHO) as a Lewis base (LB) and a bulky organoaluminum compound as a Lewis acid (LA) to construct a Lewis pair (LP), which can rapidly prepare a series of diblock copolymers (di-BCPs), poly(trifluoroethyl methacrylate)-*b*-poly(heptadecafluorodecyl methacrylate) (PTFEMA-*b*-PHDFDMA), through the sequential monomer addition method. The liquid-crystalline characteristics of PHDFDMA, in combination with Lewis pair polymerization-induced self-assembly (LP-PISA) strategy, enable ultrafast, one-pot synthesis of di-BCPs with fiber morphologies (diameters = 11.7–25.1 nm) across a wide experimental window and solid contents (up to 20% w/w). These di-BCPs are structurally characterized by ¹H nuclear magnetic resonance spectroscopy, differential scanning calorimetry, and small-angle X-ray scattering and morphologically analyzed by transmission electron microscopy. This LP-PISA strategy provides the possibility of achieving the desired morphology with well-defined structure and purity through precise control over the evolution parameters.



INTRODUCTION

Being a powerful tool for synthesis of nanomaterials, self-assembled amphiphilic AB-typed diblock copolymers (di-BCPs) have received intense attention due to their wide application potentials in biomedicine, additives, nanoreactors, and more.^{1–6} Solution self-assembly is typically employed to synthesize nanoparticles by postpolymerization in highly dilute solutions, thereby greatly restricting its practical application.^{7,8} Polymerization-induced self-assembly (PISA) was developed to simultaneously achieve both the amphiphilic copolymers and self-assemblies at high solid contents (up to 50 wt %).^{9–12} Significant progress has been made in the application of various living polymerization techniques to PISA.^{13–20} A full range of morphologies (spheres, fibers/worms, vesicles, and lamellae) could be obtained through PISA by adjusting the length of solvophilic and solvophobic blocks, solid contents, and solvents.^{21–27} Among these, fiber morphology is of particular interest due to their large surface area and anisotropic structure.^{28–32} This makes them highly applicable in various fields such as drug delivery, Pickering emulsions, super flocculants, organic photovoltaic devices, and molecular separations.^{33–37} Crystallization-driven self-assembly (CDSA) is a powerful method for synthesizing fiber morphologies through the crystallization of hydrophobic blocks with controlled lengths. However, this method typically requires highly dilute concentrations and multistep synthetic procedures.^{7,31,38–41} Introducing supramolecular structures to the

polymer chains or using liquid-crystalline (LC) polymers as hydrophobic blocks can enable the synthesis of fiber morphologies under high concentrations, but these approaches require purification processes and long reaction times, thus impairing the synthetic efficiency of PISA.^{29,37} Moreover, the experimental windows for synthesis of fibers are typically narrow, so it remains a great challenge to synthesize morphologically pure nanofibers with varying lengths and diameters.^{42,43} Therefore, it is highly desirable to develop a simple and efficient PISA strategy for fiber morphology synthesis.

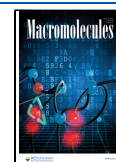
Being a newly emerged and powerful polymerization strategy, Lewis pair polymerization (LPP) has accomplished many excellent examples in polymer synthesis. These include living polymerization of various types of monomers, chemo- and regioselective polymerization of divinyl monomers, precise synthesis of polymers with controlled molecular weight (up to 2197 kg/mol), flexible regulation of the monomer sequence (up to 63-block numbers) and different topologies (linear, star,

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Scheme 1. (a) Structures of Lewis Acids and Lewis Bases. (b) Synthesis of Amphiphilic PTFEMA-*b*-PHDFDMA di-BCPs by LP-PISA

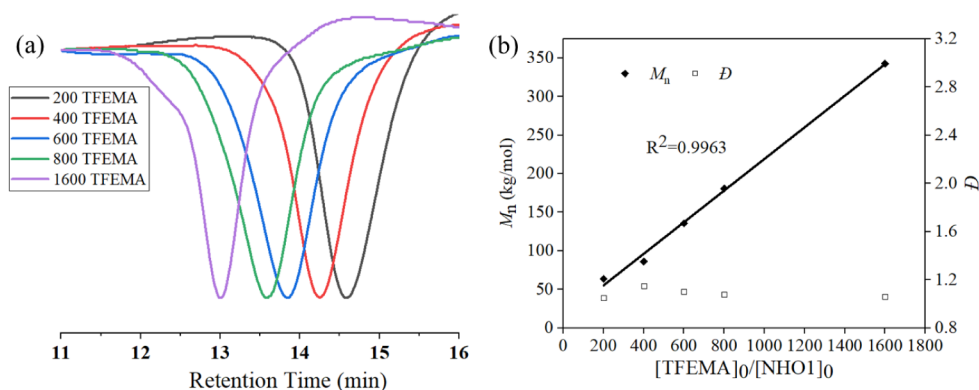
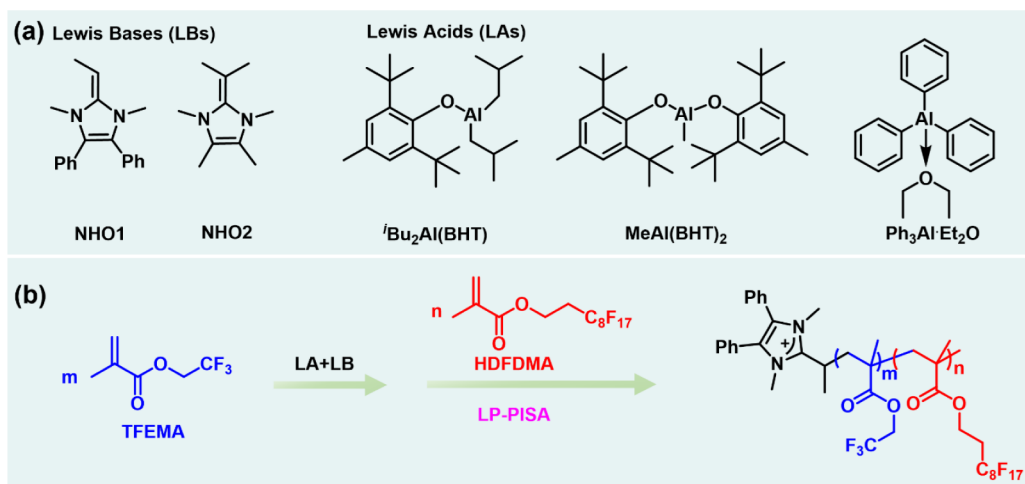


Figure 1. (a) GPC traces of PTFEMA samples prepared from polymerization performed at various $[\text{TFEMA}]_0/[\text{NHO1}]_0$ ratios at RT. Conditions: $[\text{TFEMA}]_0/[\text{NHO1}]_0/[\text{MeAl}(\text{BHT})_2]_0 = 200:1:2, 400:1:2, 600:1:2, 800:1:2$, and $1600:1:2$ and $[\text{TFEMA}]_0 = 1$ M. The negative GPC signal is due to the lower refractive index of fluoropolymer than that of the eluent. (b) Plots of M_n and $\bar{D}$ for the PTFEMA vs $[\text{TFEMA}]_0/[\text{NHO1}]_0$ ratio.

and cyclic).^{44–58} Recently, we employed LPP to obtain di-BCPs with different morphologies, which prompted us to address the above-described issues related to fiber synthesis through LP-PISA.⁵⁹ Thanks to the LC characteristics of poly(heptadecafluorodecyl methacrylate) (PHDFDMA) and excellent control of LPP, we successfully prepared amphiphilic PTFEMA-*b*-PHDFDMA di-BCPs with nanofiber morphology over a broad experimental window through an ultrafast, one-pot, two-step process. The diameters of the fibers can be tailored in the range of 11.7–25.1 nm, simply by adjusting the molar mass of the blocks and solid contents. The LC ordering of PHDFDMA was also characterized by differential scanning calorimetry (DSC) and small-angle X-ray scattering (SAXS).

RESULTS AND DISCUSSION

TFEMA Homopolymerization. Fluorinated polymers exhibit superior properties compared to that of the unfluorinated ones, including high hydrophobicity, excellent chemical and thermal properties, tunable lipophilicity, low refractive index, and low surface energy.^{60,61} PISA relies on the chain extension of a living block soluble in a selective solvent with a second monomer to generate a nonsoluble block. Due to the poor solubility of fluorinated polymers in common organic solvents, it can serve as the second monomer to generate a nonsoluble block, thereby expanding the monomer

library of PISA. Additionally, in order to synthesize amphiphilic block copolymers via PISA, it is a requisite to first obtain a soluble living block. Therefore, semifluorinated methacrylates, trifluoroethyl methacrylate (TFEMA), and heptadecafluorodecyl methacrylate (HDFDMA) were selected for PISA to maintain the balance between the solubility of monomers and polymers in solvents. First, two strong nucleophilic *N*-heterocyclic olefins (NHOs) (NHO1 and NHO2) and three organoaluminum compounds with different steric hindrance and Lewis acidity (Scheme 1a) were employed as Lewis bases (LBs) and Lewis acids (LAs) for the polymerization of TFEMA, respectively, due to their demonstrated excellent activity in (meth)acrylate polymerization.^{55,59,62} Control experiments indicated that NHO1 obtained only 4% conversion of 200 equiv of TFEMA in fluorobenzene (FB) in up to 24 h, whereas NHO2 can convert 77% of monomers to polymer within 150 min (runs 1 and 2, Table S1). The effectiveness of LAs was also examined for the polymerization of TFEMA. 21% monomer conversion was obtained by $t\text{Bu}_2\text{Al}(\text{BHT})$ alone and 7% conversion was obtained by $\text{Ph}_3\text{AlEt}_2\text{O}$, whereas $\text{MeAl}(\text{BHT})_2$ showed no activity for the polymerization of TFEMA in 24 h (runs 3–5, Table S1). To avoid the background polymerization initiated by LB or LA itself, NHO1 and $\text{MeAl}(\text{BHT})_2$ were selected and a Lewis pair was constructed to examine the polymerization of

Table 1. PISA Results^a

run	LB/LA/TFEMA/HDFDMA	solid content (%)	time (min)	conv. ^b (%)	morphology ^c	diameter (nm) ^c
1	1/2/100/25	15	1/1	100/100	fiber	11.2
2	1/2/100/40	15	2/3	100/100	fiber	13.8
3	1/2/100/25	20	1/1	100/100	fiber	11.7
4	1/2/100/40	20	1/2	100/100	fiber	21.8
5	1/2/100/25	5	5/5	100/100	fiber	13.7
6	1/2/100/50	5	5/6	100/100	fiber	22.5
7	1/2/200/25	15	3/4	100/100	fiber	13
8	1/2/200/50	15	3/4	100/100	fiber	16.5
9	1/2/200/75	15	3/7	100/100	fiber	18.3
10	1/2/200/25	5	7/3	100/100	fiber	15.2
11	1/2/200/50	5	14/26	100/100	fiber	17.1
12	1/2/200/75	5	20/20	100/94	fiber	20.9
13	1/2/200/75	10	6/9	100/100	fiber	15.8
14	1/2/200/50	20	6/9	100/100	fiber	14.5
15	1/2/200/75	20	2/5	100/100	fiber	18.4
16	1/2/300/50	15	5/4	100/100	fusiform+short fiber	14.5
17	1/2/300/75	15	6/3	100/79	fusiform+short fiber	17.3
18	1/2/300/125	15	7/18	99/100	fusiform+short fiber	28.5
19	1/2/300/100	10	10/20	100/100	fusiform	62.2/24.5 ^d
20	1/2/300/75	20	4/3	100/100	long fiber+short fiber+fusiform	18.1
21	1/2/600/100	15	20/55	100/100	fusiform	60.7/25.5 ^d
22	1/2/600/200	15	35/43	100/80	fusiform	71.9/39.2 ^d

^aCondition: carried out at ambient temperature (~25 °C) in FB at solid contents of 5–20 wt %, where solid content = $[m(\text{catalysts}) + m(\text{monomers})]/[m(\text{catalysts}) + m(\text{monomers}) + m(\text{solvent})] \times 100\%$. ^bMonomer conversion was measured by ¹H NMR spectroscopy.

^cMorphologies were determined by TEM. ^dThe length and width of the fusiform.

TFEMA. Quantitative monomer conversion was obtained in 1 min for the polymerization performed in a 200:1:2 [TFEMA]₀/[NHO1]₀/[MeAl(BHT)₂]₀ ratio (run 6, Table S1), furnishing PTFEMA with $M_n = 63.5$ kg/mol and $\bar{D} = 1.05$. Increasing the [TFEMA]₀/[NHO1]₀ ratio from 200 to 1600, full monomer consumptions can be obtained for all of the investigated ratios, producing polymers with predicted M_n and narrow $\bar{D}$ (runs 6–10, Table S1). GPC traces showed a gradual shift to higher molar mass regions, while maintaining narrow and unimodal distribution (Figure 1a). The M_n values of the resulting PTFEMA increased linearly ($R^2 = 0.996$) from 63.5 to 343 kg/mol as the monomer-to-LP ratio increased from 200 to 1600, while the $\bar{D}$ values were kept in a narrow range between 1.05 and 1.15 (Figure 1b). Furthermore, a plot of PTFEMA M_n vs monomer conversion at a fixed [TFEMA]₀/[NHO1]₀/[MeAl(BHT)₂]₀ ratio of 400:1:2 also gave a straight line ($R^2 = 0.996$), which was coupled with the small $\bar{D}$ values (Figure S2). The analysis of the low-MW PTFEMA sample with matrix-assisted laser desorption/ionization time-of-flight mass spectroscopy (MALDI-TOF MS) revealed that the polymer produced by NHO1/MeAl(BHT)₂ is a linear polymer chain capped with NHO1/H groups (Figures S3 and S4). In contrast, a few series of molecular ion peaks were detected for the PTFEMA produced by NHO alone, where the major series corresponds to linear PTFEMA capped with NHO2 and cyclic β -ketoester or δ -valerolactone chain ends resulting from backbiting side reactions (Figures S5 and S6), indicating that the polymerization by NHO2 alone is not living and controlled.

Our previous work revealed that the reaction of NHO1 and MeAl(BHT)₂ formed a pure classic Lewis adduct (CLA).⁵⁵ While the stoichiometric reaction performed in a 1:1:1 NHO1:MeAl(BHT)₂:TFEMA ratio produced zwitterionic enolaluminate intermediates as two isomers (Figure S7), this

polymerization adopted a bimolecular activation mechanism. In order to verify the livingness feature of this polymerization, chain extension experiments were performed by adding the second batch of monomers after completion of the first batch of monomers without quenching. GPC traces of the resulting polymers shifted to the higher-molar-mass region and showed a narrow and unimodal molecular weight distribution (MWD) (Figure S1). The third batch of monomers can still be fully consumed. The M_n values of the polymers produced from the chain-extension experiments increased from 63.5 to 88.5 to 114 kg/mol, respectively (runs 1–3, Table S2). All of these results clearly demonstrated the excellent controllability of NHO1/MeAl(BHT)₂ LP over the TFEMA polymerization. Therefore, NHO1/MeAl(BHT)₂ was selected for the following investigations.

Dispersion Polymerization of HDFDMA. After obtaining this soluble, living PTFEMA polymer chain, we turned our focus to its application in PISA. Due to the LC characteristics and insolubility of PHDFDMA in FB, HDFDMA was utilized as core-forming monomer to copolymerize with TFEMA through a sequential monomer addition method, with 5–20% solid contents (Scheme 1b). First, with a fixed degree of polymerization (DP) of PTFEMA at 100 and solid content at 15%, after completing TFEMA within 1 min, HDFDMA was added at DP = 25, and full monomer conversion was obtained within 1 min as revealed by ¹H NMR spectroscopy (run 1, Table 1). Due to their poor solubility in common organic solvents, the resulting copolymers were not analyzed by GPC. By using 1,3-bis(trifluoromethyl)benzene as a cosolvent (1,3-bis(trifluoromethyl)benzene:CDCl₃ = 1:5 v/v), the copolymers were characterized by ¹H NMR spectroscopy, which confirmed that the composition of the produced copolymers is consistent with that of the target PTFEMA₁₀₀-*b*-PHDFDMA₂₅ di-BCPs (the subscripts represent the DP of each block, Figure

S8). Moreover, transmission electron microscopy (TEM) images revealed the formation of pure nanofibers with an average diameter of 11.2 nm (Figure 2a). With the DP of

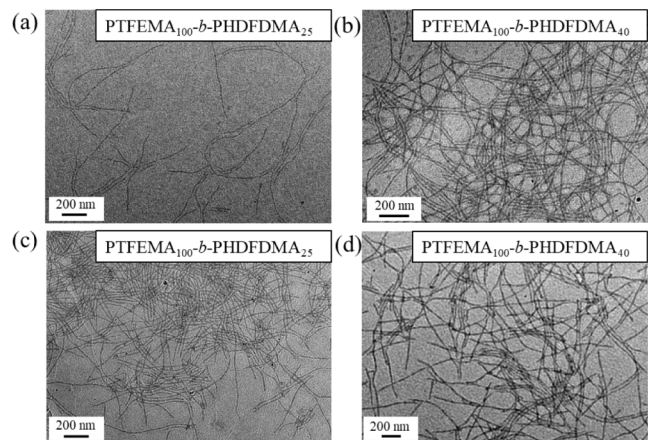


Figure 2. TEM images for various di-BCP nanoparticles prepared by LP-PISA: (a) PTFEMA₁₀₀-*b*-PHDFDMA₂₅ at 15 wt % solid content; (b) PTFEMA₁₀₀-*b*-PHDFDMA₄₀ at 15 wt % solid content; (c) PTFEMA₁₀₀-*b*-PHDFDMA₂₅ at 20 wt % solid content; (d) PTFEMA₁₀₀-*b*-PHDFDMA₄₀ at 20 wt % solid content.

PHDFDMA increasing to 40, pure nanofibers could still be obtained within 5 min and the average diameters of nanofibers increased to 13.8 nm (Figure 2b). To the best of our knowledge, it is the first time that pure nanofibers have been prepared in such a short time using the PISA strategy.^{63,64}

Solid content is also found to play an essential role in affecting the morphologies of the assemblies. Therefore, we further investigated the effects of solid content on morphologies and found that with the solid contents ranging between 5% and 20%, pure nanofibers were obtained as the only morphology, with average diameters in the range of 11.7–22.5 nm (runs 3–6, Table 1, Figures 2c,d and S13,14). To examine the capability of LP-PISA in synthesizing a pure nanofiber phase with a wider experimental window, we fixed the solvophilic PTFEMA at DP = 200 and varied the DP of PHDFDMA from 25 to 50, and 75, respectively. For the copolymerization performed with PHDFDMA at DP = 25, short nanofibers were obtained with an average diameter = 13 nm in 7 min (run 7, Table 1, and Figure S15), while the average diameter increased to 16.5 and 18.3 nm, as the DP of

PHDFDMA increased to 50 and 75, respectively (runs 8 and 9, Table 1, and Figure S16). TEM images showed that the nanofibers could be produced by the polymerization performed with solid contents ranging between 5% and 20% (runs 10–15, Table 1, Figures S17–22). These results obtained at different DPs and solid contents showed that the experimental window for the synthesis of pure nanofibers is very wide, highlighting the robustness of this LP-PISA strategy.

As the LC ordering is the important driving force for the formation of nanofibers in a large experimental window, SAXS was employed to identify the phase structures of a series of di-BCP samples. Both PTFEMA₂₀₀-*b*-PHDFDMA₂₅ and PTFEMA₂₀₀-*b*-PHDFDMA₅₀ exhibited a principle scattering peak $q = 1.99 \text{ nm}^{-1}$ and a second order peak at $q = 3.99 \text{ nm}^{-1}$, where the position of the second order peak was two times that of the principle scattering peak (Figure 3a). It is also noted that the interlayer spacing of lamellae calculated from the position of the principal scattering peak, q (as $d = 2\pi/q$), is 3.16 nm, which is almost twice the length of the extend side-chain length of PHDFDMA (1.59 nm) (Figure S26). In combination with the fact that there is no observation of scattering peak in the SAXS spectrum of PTFEMA, the LC phase of the PHDFDMA in the assemblies should be the smectic A (SmA) phase. Moreover, DSC was utilized to determine the SmA-to-isotropic phase transition temperature (T_c) of the above-described three samples (Figure 3b). It turned out that there is no T_c observed for PTFEMA homopolymer, while it is 83 °C for PTFEMA₂₀₀-*b*-PHDFDMA₂₅ and 88 °C for PTFEMA₂₀₀-*b*-PHDFDMA₅₀, which further confirmed that the stability was increased as the length of PHDFDMA increased.

Study of the Morphology Evolution. To understand the morphology evolution of the nanofibers and identify the structures of important intermediates, we further investigated the influencing effects of PTFEMA with longer length on PISA. As expected, with a fixed 300 DP of PTFEMA and 15 wt % solid content, copolymerization performed with the DP of PHDFDMA at 50 obtained PTFEMA₃₀₀-*b*-PHDFDMA₅₀ assemblies as a mixture of fusiform micelles and short fiber-like micelles (run 16, Table 1, Figure 4a). These short fiber-like micelles might form by fusing fusiform micelles into dimers, trimers, and longer structures. Increasing the DP of PHDFDMA from 50 to 75 or 125 resulted in mixed morphologies, with both the length and width of the fusiform micelles increasing (Figures 4b and S23, runs 17 and 18, Table 1). At a 10 wt % solid content, pure fusiform micelles were obtained for PTFEMA₃₀₀-*b*-PHDFDMA₁₀₀ assemblies (length

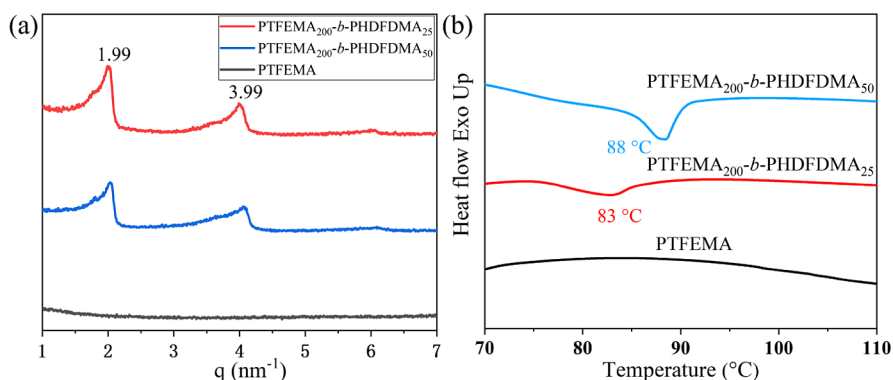


Figure 3. (a) SAXS spectra of PTFEMA, PTFEMA₂₀₀-*b*-PHDFDMA₂₅ and PTFEMA₂₀₀-*b*-PHDFDMA₅₀; (b) DSC curves of PTFEMA, PTFEMA₂₀₀-*b*-PHDFDMA₂₅ and PTFEMA₂₀₀-*b*-PHDFDMA₅₀.

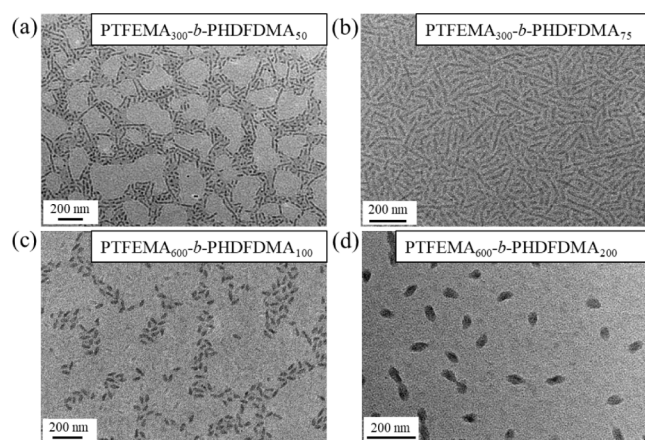


Figure 4. TEM images for various di-BCP nanoparticles prepared at 15 wt % solid contents at RT: (a) PTFEMA₃₀₀-*b*-PHDFDMA₅₀; (b) PTFEMA₃₀₀-*b*-PHDFDMA₇₅; (c) PTFEMA₆₀₀-*b*-PHDFDMA₁₀₀; (d) PTFEMA₆₀₀-*b*-PHDFDMA₂₀₀.

= 62.2 nm, width = 24.5 nm, run 19, Table 1, and Figure S24). However, at 20% solid content, PTFEMA₃₀₀-*b*-PHDFDMA₇₅ assemblies were obtained with mixed morphologies including long fibers, short fibers, and fusiform micelles (run 20, Table 1, and Figure S25), which is consistent with the previous results suggesting that high solid content may be beneficial for the formation of pure high-order morphologies (such as fiber/worms or vesicles).^{21,42,65} With the DP of PTFEMA set at 600, only pure fusiform micelles were formed regardless of whether the DP of PHDFDMA was 100 or 200 (length = 60.7 or 71.9 nm, width = 25.5 or 39.2 nm, Figure 4c,d, runs 21 and 22, Table 1), confirming that nanofibers originate from the fusion of fusiform micelles. Thanks to the excellent controllability, high synthetic efficiency, and wide experimental window of this LP system, polymeric nanofibers with adjustable width and length can be prepared. These fluorinated nanofibers are expected to show application potentials in the additives of epoxy resins to enhance the fracture resistance, thermoresponsive materials, and ¹⁹F magnetic resonance image agent.^{34,66,67}

CONCLUSION

In summary, we have developed a highly efficient NHO-based LP to realize living/controlled polymerization of semi-fluorinated methacrylates, TFEMA. Using a sequential monomer addition method, PTFEMA-*b*-PHDFDMA di-BCPs were synthesized with a well-defined structure in a one-pot two-step process. The smectic ordering of solvophobic PHDFDMA, coupled with the LP-PISA strategy, enabled the ultrafast synthesis of nanofiber morphologies (11.7–25.1 nm) across a wide experimental window of 5–20% solid contents, achieved by changing the TFEMA to HDFDMA ratio. The well-defined LC ordering of PTFEMA-*b*-PHDFDMA di-BCPs were also confirmed by SAXS and DSC analysis. Investigations into morphology evolution indicated that the nanofibers are formed through the fusion of fusiform micelles. The LP-PISA strategy, without the requirement for isolation and purification of macroinitiators, exhibited significantly enhanced synthetic efficiency, thereby paving the way for large-scale production of assemblies in the future.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.macromol.4c00768>.

General experimental methods, synthesis and characterization details, general polymerization methods and polymer characterization, some other polymerization results, and collected NMR (¹H, ¹³C) spectra (PDF)

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Author Contributions

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Notes

The authors declare no competing financial interest.

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